

1 **European Network for Optimization of Veterinary Antimicrobial Therapy (ENOVAT) Guidelines**
2 **for Antimicrobial Use in Canine Acute Diarrhoea**

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39 **Abstract**

40 Acute diarrhoea is a common presentation in dogs, and a common reason for antimicrobial
41 prescription and nutraceutical use. This evidence-based guideline provides recommendations for
42 antimicrobial and nutraceutical treatment of canine acute diarrhoea (CAD). A multidisciplinary panel
43 developed the recommendations by adhering to the Grading of Recommendations Assessment,
44 Development and Evaluation (GRADE) framework. The opinions of stakeholders (general veterinary
45 practitioners and dog owners) were collected and incorporated to ensure the applicability of this
46 guideline. Four strong recommendations informed by high certainty evidence, and three conditional
47 recommendations informed by very low or low certainty evidence, were drafted by the panel, along with
48 an ungraded section on diagnostic work-up of dogs with acute diarrhoea. The ENOVAT guidelines
49 initiative encourages national or regional guideline makers to use the evidence presented in this
50 document, and the supporting systematic review, to draft national or local guidance documents.

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53 **Keywords**

54 **Antimicrobial stewardship; antibiotics; enteritis; evidence based; GRADE**

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<p>Recommendation 1 In dogs with mild disease and acute non-hemorrhagic diarrhoea (dogs in good general condition, with no signs of dehydration or systemic illness), we recommend against treatment with antimicrobials. <i>Strong recommendation, high-certainty evidence.</i></p>
<p>Recommendation 2 In dogs with mild disease and acute diarrhoea with hematochezia (dogs in good general condition, with no signs of dehydration or systemic illness), we recommend against treatment with antimicrobials. <i>Strong recommendation, high-certainty evidence.</i></p>
<p>Recommendation 3 In dogs with acute non-hemorrhagic diarrhoea, and moderate disease (dogs with impaired general condition and varying degrees of dehydration/hypovolemia. Dogs may have signs of systemic disease related to the deficit of body fluids, that will resolve with adequate fluid therapy), we recommend against treatment with antimicrobials. <i>Strong recommendation, high-certainty evidence.</i></p>
<p>Recommendation 4 In dogs with acute hemorrhagic diarrhoea, and moderate disease (dogs with impaired general condition and varying degrees of dehydration/hypovolemia. Dogs may have signs of systemic disease related to the deficit of body fluids that will resolve with adequate fluid therapy), we recommend against treatment with antimicrobials. <i>Strong recommendation, high-certainty evidence. Remarks:</i> Dogs with laboratory values indicative of severe or overwhelming inflammation, such as severe neutrophilia ($> 25 \times 10^9/L$), neutropenia and/or degenerative left-shift, represent an exception.</p>
<p>Recommendation 5 In dogs with hemorrhagic and non-hemorrhagic diarrhoea, and severe disease (dogs with impaired general condition and varying degrees of dehydration/hypovolemia, and signs of systemic disease despite adequate fluid therapy), we suggest systemic treatment with antimicrobials. <i>Conditional recommendation, very low-certainty evidence.</i></p>
<p>Recommendation 6 In dogs with severe disease, we suggest parenteral (intravenous or intramuscular) administration of antimicrobials that are expected to be effective for treatment of bacterial translocation and bacteraemia or sepsis. Drug choice depends on how critical the clinical status of the dog is, as well as regional prevalence of antimicrobial resistance (AMR) and drug availability. In dogs with non-critical illness (Table 1) living in a region with low AMR prevalence, we suggest ampicillin or alternatively amoxicillin-clavulanic acid or trimethoprim/sulfonamides as first line drugs. In dogs with critical illness (Table 1) or where antimicrobial resistance is more likely (e.g. based on geographic trends or the patient's antimicrobial exposure history) we suggest administration of a four-quadrant protocol providing gram positive, gram negative, aerobic and anaerobic coverage (Table 5). Dogs with non-critical illness that do not respond to first line antimicrobials and supportive care should also receive this protocol. <i>Conditional recommendation, very low-certainty evidence/expert opinion. Level of agreement 100 %</i> *Antimicrobial drug combinations with four-quadrant spectrum (aerobic, anaerobic, gram positive and gram negative spectrum) include aminopenicillins or clindamycin combined with fluoroquinolones or aminoglycosides (gentamicin, amikacin).</p>
<p>Recommendation 7 The duration of antimicrobial treatment is dependent on the treatment response and the panel suggests daily assessment of animals while hospitalized. Antimicrobial therapy should not extend beyond clinical resolution. For the majority of dogs, treatment of 3-7 days is likely adequate to obtain clinical resolution <i>Conditional recommendation, very low-certainty evidence. Level of agreement 100%</i></p>
<p>Recommendation 8 In dogs with acute diarrhoea we do not recommend either for, or, against use of probiotics. <i>The trade-offs are closely balanced. Moderate certainty evidence.</i></p>

68 **Introduction**

69 Acute diarrhoea in dogs is a common presenting complaint in veterinary practice ((Jones et al., 2014).
70 The vast majority of dogs with acute diarrhoea have mild and self-limiting disease (Hubbard et al., 2007),
71 while a small proportion of dogs become more profoundly sick and require intravenous fluid support and
72 hospitalization (Singleton et al., 2019). A study of over 3000 dogs with acute diarrhoea presented to
73 primary practice showed that 84% of consults had mild clinical signs, 15 % had moderate clinical signs,
74 and less than 1% had severe clinical signs, as defined by the attending veterinarian (Singleton et al.,
75 2019). Only 2.3% of all dogs were admitted and 0.2% were referred to secondary practice in the same
76 study. While the aetiology of acute diarrhoea often remains unknown, the prognosis in most cases is
77 excellent. Most cases resolve within one week (Hubbard et al., 2007) and fatalities are rare, with an all-
78 cause mortality/euthanasia in hospitalized dogs of approximately 2– 4 % (Mortier et al., 2015; Dupont et
79 al., 2021). Despite the mild biological course of disease and favorable prognosis, acute diarrhoea remains
80 one of the more common indications for antimicrobial use in dogs (De Briyne et al., 2013). It has been
81 documented that 50-65% of dogs with acute diarrhoea are prescribed antimicrobials (Jones et al., 2014;
82 Singleton et al., 2019; Lutz et al., 2020). According to a UK study, metronidazole is most frequently
83 administered drug followed by amoxicillin-clavulanic acid (Singleton et al., 2019). Antimicrobial
84 resistance (AMR) is one of our times most pressing health problems, it affects humans and animals alike,
85 and is mainly driven by the selection pressure created by antibiotic usage (WHO, 2024). Canine acute
86 diarrhea represents a highly common condition associated with inappropriately high antimicrobial
87 prescription rates, and as such, is of high priority for antimicrobial stewardship in companion animal
88 practice. Presently there are no international antimicrobial use guidelines available for treatment of acute
89 diarrhea in dogs.

90 **Scope and purpose**

91 The purpose of this document is to provide guidance on antimicrobial use in dogs with acute diarrhoea,
92 based on the best available evidence and transparent reasoning. The target audience is veterinary
93 practitioners managing dogs with acute diarrhoea, in either out-patient or hospital settings. The guideline
94 is intended to help practitioners direct antimicrobial treatment towards those dogs that are most likely to
95 benefit from it, while reducing unnecessary use in the remaining dogs. As with all guidelines, this
96 document is not intended to be a substitute for good clinical judgement, and recommendations should not
97 be viewed as diktats. Even strong recommendations may not apply to all dogs in all circumstances.

98 The recommendations in this guideline are informed by the systematic review previously published by the
99 group (Scahill et al., 2024). The ENOVAT guidelines initiative encourages national or regional guideline
100 makers to use the evidence presented in this document, and the supporting systematic review, to draft

101 national or local guidance documents. Translation and dissemination of ENOVAT guidance documents is
102 encouraged.

103 This guideline is produced in collaboration with the European Society of Clinical Microbiology and
104 Infectious Disease (ESCMID) Study Group for Veterinary Microbiology (ESGVM).

105 **Methods**

106 This guideline was produced following the ENOVAT operating procedure (ENOVAT, 2024). The
107 Grading of Recommendations, Assessment, Development and Evaluation (GRADE) approach was used
108 to assess the certainty of the evidence and draft recommendations (Guyatt et al., 2008).

109 *Composition of the Guidelines Drafting Group*

110 The guidelines panel was established in 2020, and is composed of 18 members representing the veterinary
111 fields of gastroenterology (MW, SU, CRB, KA), internal medicine, (LRJ, FA, EL, CA, CP), infectious
112 diseases (SW, KS), general medicine (TB), microbiology (LG), pharmacology (AF), epidemiology (MB,
113 DS) and public health (UW). One panel member (FF) represents the field of guidelines methodology in
114 human medicine. The work was chaired by an oversight committee (LRJ, DS) and a methodology
115 taskforce (KS, MW, CP, MB, FF) was established as a subset of the group. Two members of the
116 methodology taskforce were non-voting members (MB, FF).

117 *Conflict of interest*

118 This article is based upon work from the COST Action European Network for Optimization of Veterinary
119 Antimicrobial Treatment (CA18217), supported by COST (European Cooperation in Science and
120 Technology). The panel members did not have any substantial conflicts of interest at the time of drafting
121 recommendations. However, it should be noted that most of the panel members are involved in
122 antimicrobial stewardship activities. Two panel members (MW and SU) were authors of trials included in
123 the systematic reviews and did not participate in the risk of bias assessment or any other individual
124 quality assessments for these publications.

125 *Generation of Guidelines content and involvement of veterinary practitioners and dog owners*

126 An overview of the guidelines process is depicted in Figure 1. In brief, the content of the guidelines and
127 the clinical questions were generated by the panel in an iterative process involving electronic Delphi
128 questionnaires and on-line meetings. The panel defined the target population as dogs with acute (less than
129 7 days duration) diarrhoea, regardless of aetiology, and categorized this population into three sub-
130 populations of dogs depending on the severity of their clinical state. Each sub-population was further sub-

131 grouped, based on the presence or absence of blood in the stools. Three clinical questions concerning the
132 effect, choice and duration of antimicrobial therapy, were selected for systematic reviews. Furthermore,
133 three clinical questions concerning the effect of nutraceuticals were selected for systematic review, of
134 which only the question on probiotics was included in the guidelines.

135 Clinical questions were phrased using the Population Intervention Comparator Outcome (PICO) format.
136 To ensure the relevance of the guidelines content, and integrated the perspectives of guideline end-users,
137 panel members conducted structured interviews with veterinary practitioners (n=41) and dog-owners
138 (n=33) from across Europe and Israel. From this process, five outcomes (duration of diarrhoea,
139 progression of disease, duration of hospitalization, mortality and adverse effects) were prioritized for
140 evaluation. Outcomes were classified as critical if deemed so by the majority of either the veterinary
141 practitioners, dog owners and/or panel members.

142 To evaluate the effect of treatment, thresholds for clinically relevant treatment effects were established for
143 all outcomes. Thresholds for a relevant reduction in the duration of diarrhoea, and a relevant reduction in
144 the duration of hospitalization, were established prior to conducting the systematic review, and were
145 based on the opinion of the majority of interviewed veterinary practitioners, dog owners and panel
146 members. The thresholds for a clinically relevant effect of treatment on the risk of mortality, and the risk
147 of disease progression, were established after conducting the systematic review, following GRADE's
148 updated guidance of the imprecision domain (Zeng *et al.*, 2022). These thresholds were derived by
149 surveying a different group of veterinary practitioners (n=23) and panel members from the clinical field
150 (n=11) and calculating the 25-percentile value of the risk-effects selected by the survey participants.
151 Outcomes and thresholds for a clinically relevant treatment effect are listed in Table 1. Subgrouping of
152 dogs are described in table 2.

153 Table 1. Critical outcomes and treatment effect thresholds in dogs with acute diarrhea

Outcome (subgroup)	Threshold for a clinically relevant effect of treatment
Duration of diarrhea	At least 1 day reduction
Duration of hospitalization (dogs with moderate and severe disease)	At least 1 day reduction
Mortality (dogs with severe disease)	3 % risk increase/decrease
Progression of disease (dogs with mild disease)	30 % risk increase/decrease
Progression of disease (dogs with moderate – severe disease)	10 % risk increase/decrease

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155 *Systematic review and judging the certainty of evidence*

156 The systematic reviews, meta-analyses (MA), and evidence assessment were conducted by the
157 methodology taskforce and oversight committee. The results of the systematic reviews, and a description
158 of the methods applied, are available in the supporting systematic review (Scahill et al., 2024). In brief,
159 the certainty of evidence was assessed for each outcome using the GRADE methodology, and was based
160 on the risk of bias, imprecision, indirectness, inconsistency and publication bias (Guyatt et al., 2008). The
161 partially contextualized approach was used to assess imprecision for separate outcomes (Zeng et al.,
162 2022). The certainty of the body of evidence was based on the certainty of evidence of the outcomes
163 deemed critical, and could not be graded higher than the critical outcome with the lowest certainty.

164 *Generation of recommendations*

165 Recommendations were drafted by the panel in May 2022 in a face-to-face hybrid meeting in
166 Copenhagen. Prior to the meeting, panel members were presented with a video summary of the systematic
167 review and meta-analyses, as well as a written evidence summary report prepared by members of the
168 methodology taskforce (KS, MW). Panel members were also provided with a narrative summary of the
169 harmful effects of antimicrobial therapy on the canine gastrointestinal residual flora (MW, SU, LG), and a
170 summary of the stakeholder interviews (LRJ, CP). Finally, panel members were provided with links and
171 asked to familiarize themselves with video material from the McMaster University on the guidelines
172 formation process following the GRADE approach. Drafting of recommendations followed the GRADE
173 Evidence to Decision (EtD) framework, and for each recommendation the following factors were
174 discussed: certainty of the overall evidence, the balance of desirable and undesirable effects, preferences
175 and values of dog-owners and veterinary practitioners, equity, acceptability and feasibility (Alonso-
176 Coello et al., 2016). The panel defined consensus as 80% agreement prior to drafting recommendations.
177 Agreement was calculated based on the 16 voting members. The panel drafted four strong and three
178 conditional recommendations. Strong recommendations were informed by moderate or high certainty
179 evidence, conditional recommendations were informed by low or very low certainty evidence. The
180 definitions of certainty and the implications of strong and conditional recommendations are described in
181 Table 3. Two recommendations (6 and 7) were elaborated and modified after the meeting and subjected to
182 two more processes of agreement. All recommendations received 100 % agreement.

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185 *Generation of the diagnostic (ungraded) section.*

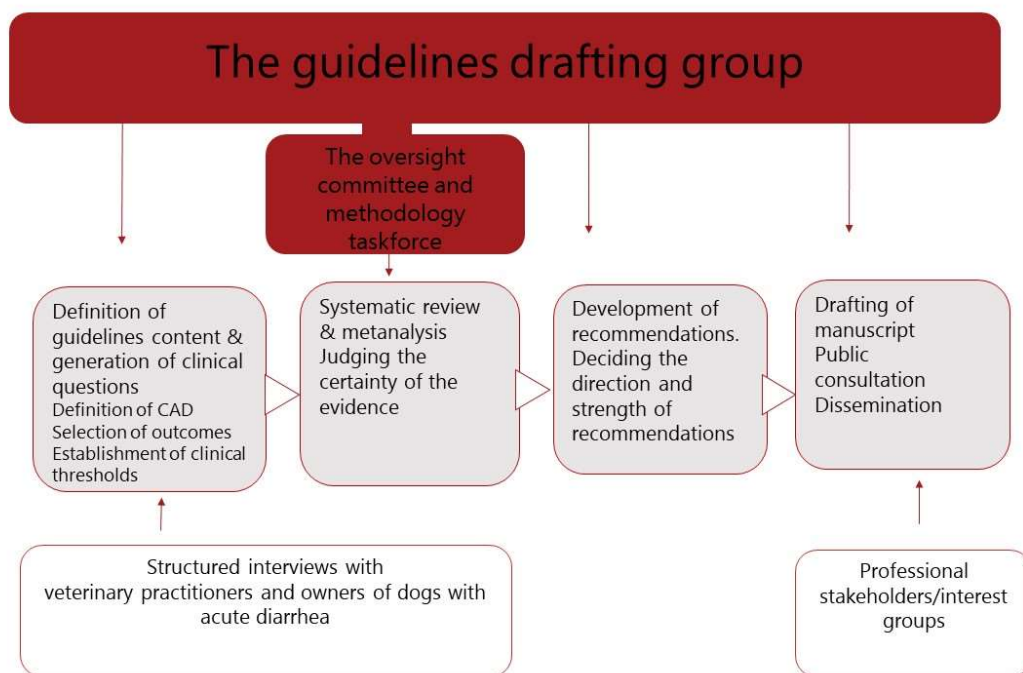
186 The diagnostic section was generated by an iterative process involving several Delphi rounds and a final
187 approval of considerations by the voting panel members.

188 *Consultation phase*

189 Guidelines were available on the ENOVAT website from 26/02/2024 to 26/03/2024 for public
190 consultation (ENOVAT, 2024). The public consultation phase was announced by the ENOVAT
191 newsletter and members from ESGVM, ENOVAT and the European Society of Comparative
192 Gastroenterology (ESCGE) were contacted by email/newsletter and encouraged to participate.

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197 Fig 1. Overview of the guidelines process.

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203 Table 2. Sub-populations of dogs with acute diarrhea.

Sub-population	Presence/absence of blood in the stools	Definition
Mild disease	Non-hemorrhagic diarrhea Hematochezia.	Dogs with mild disease are bright, alert and responsive. They have no clinical signs of dehydration or hypovolemia and there is absence of fever. These dogs are managed as out-patients.
Moderate disease	Non-hemorrhagic Hemorrhagic	Dogs with moderate disease have mild to moderately depressed mental status, and are dehydrated or hypovolemic. Dogs in this category may present with signs of systemic disease, typically tachycardia. When present, systemic signs are due to dehydration/hypovolemia, and resolve rapidly with adequate fluid replacement. There is absence of fever. Dogs with moderate disease warrant fluid therapy and supportive care, and are often hospitalized.
Severe disease	Non-hemorrhagic Hemorrhagic	Dogs with severe disease have moderately to severely depressed mental status and signs of dehydration or hypovolemia. Dogs with severe disease warrant fluid therapy and supportive, sometimes intensive care. These dogs are hospitalized. Dogs in this category may present in different ways: <ul style="list-style-type: none"> • Dogs with critical illness (severely depressed mental status and severe vascular compromise/shock) • Dogs with non-critical illness e.g., dogs presenting with moderate disease but systemic signs do not resolve or progress or relapse despite adequate fluid replacement; or dogs that present with or develop overwhelming inflammation (severe neutrophilia or neutropenia) or fever (>39.3)

204 *Dogs with acute diarrhea are sub-categorized according to the severity of clinical disease.*

205 *Categorization is not based on volume or frequency of diarrhea.*

206

207 Table 3. Definition of the certainty of evidence and implications of strong versus conditional
208 recommendations.

Certainty of evidence ^a	
High	The authors have a lot of confidence that the true effect is close to the estimated effect.
Moderate	The authors believe that the true effect is probably close to the estimated effect.
Low	The true effect might be markedly different from the estimated effect.
Very low	The true effect is probably markedly different from the estimated effect

Recommendations		
Implications for:	Strong Recommendation	Conditional Recommendation
Animals	Most animals in this situation would benefit from the recommended course of action and only a small proportion would not.	The majority of animals in this situation would benefit from the suggested course of action, but many would not.
Clinicians	Most animals should receive the recommended course of action.	Evidence is inadequate to make a strong recommendation, and/ or different choices might be appropriate for different animals. Be prepared to help animal owners make a decision that is consistent with their own values/preferences.
Policy makers	The recommendation can be adapted as policy in most situations.	Policy making may require substantial debate and involvement of many stakeholders. Policies are also more likely to vary between regions.

209 Modified from (Guyatt et al., 2008)

210

211 **Results**

212 **Recommendations on antimicrobial use in dogs with acute diarrhoea and mild disease**

213 **Recommendation 1**

214 In dogs with mild disease and acute non-hemorrhagic diarrhoea we recommend against treatment with
215 antimicrobials.

216 *Strong recommendation, high-certainty evidence.*

217 *Level of agreement 100%*

218

219 **Recommendation 2**

220 In dogs with mild disease and acute diarrhoea with hematochezia we recommend against treatment with
221 antimicrobials.

222 *Strong recommendation, high-certainty evidence.*

223 *Level of agreement 100%*

224 *Rationale for recommendations 1 & 2*

225 *Evidence of therapeutic effect*

226 There is high certainty evidence that antimicrobials do not confer a clinically relevant effect in dogs with
227 acute diarrhoea and mild disease, whether or not blood is present in the stools. Based on the enquiries
228 among dog owners and veterinarians, the main concern in dogs with acute diarrhoea and mild disease is
229 the duration of diarrhoea (critical outcome), and for dogs with hematochezia, the risk of disease
230 progression is also a concern (critical outcome). To investigate the effect of antimicrobials in dogs with
231 diarrhoea, we conducted a systematic review, and included outcome data from 232 dogs from six
232 randomized controlled trials in a meta-analysis (Scahill et al., 2024). Dogs with mild disease were
233 represented in four trials (Shmalberg et al., 2019; Langlois et al., 2020; Werner et al., 2020; Rudinsky et
234 al., 2022), two of which also included dogs with moderate disease and non-hemorrhagic diarrhoea
235 receiving intravenous fluid therapy as out-patients (Shmalberg et al., 2019; Langlois et al., 2020). Dogs
236 with hematochezia were represented in one study (Rudinsky et al., 2022). The remaining two trials were
237 conducted in dogs with moderate disease and hemorrhagic diarrhoea (Unterer et al., 2011; Israeloff,
238 2009). Antimicrobials investigated were metronidazole (3 studies), amoxicillin clavulanate (2 studies) or
239 a combination (1 study).

240 The mean duration of diarrhoea in dogs with acute diarrhoea ranged from 1.7 to 9.3 days in dogs
241 receiving antimicrobials and from 1.9 to 6.68 days in the control group. When looking at the pooled mean
242 difference between treated and untreated dogs, duration of diarrhoea was reduced by 0.28 days or
243 approximately 7 hours (95% CI -0.77-0.21) in dogs receiving antimicrobials. The mean difference was
244 below the 24 hours threshold for a clinically relevant reduction in the time of diarrhoea, as predefined by
245 dog-owners and veterinary practitioners, and was therefore considered trivial. Likewise, subgroup
246 analysis of the 126 non-hospitalized dogs (dogs with mild disease, and dogs with moderate disease and
247 non-hemorrhagic diarrhoea) and the 106 hospitalized dogs (dogs with moderate disease and hemorrhagic
248 diarrhoea) showed only trivial reduction in the duration of diarrhoea in response to antimicrobials. The
249 mean reductions in days of diarrhoea were 0.07 days (95% CI -1.19-1.05) and 0.38 days (95% CI -0.81-
250 0.04), respectively. No dogs with mild disease included in the systematic review suffered progression of
251 disease. The certainty of evidence for dogs with mild disease was high. The systematic review included
252 the six trials in a network meta-analysis to make an indirect comparison between metronidazole and beta-
253 lactams. Amoxicillin-clavulanic acid was marginally more efficient in shortening the duration of
254 diarrhoea (MD -0.29 days, 95% CI -2.24, 1.65) in comparison to metronidazole but the difference was
255 considered clinically trivial (below 24 hours), and did not change the overall conclusion (Scahill et al.,
256 2024).

257 *The balance between desirable and undesirable effects*

258 From the perspective of the individual dog and society, avoidance of antimicrobial use, where there is no
259 benefit of therapy, is preferred to avoid harmful effects of antimicrobial treatment (Table 4). Harmful
260 effects could include adverse drug effects, antimicrobial resistance, impacts on the gut microbiota, and
261 problems relating to drug administration (e.g., bites, disruption of the human-animal bond).

262 The six studies included in the systematic review did not report adverse effects, or exacerbation of clinical
263 signs, in association with antimicrobial administration. However, adverse effects may go undetected in
264 dogs with acute diarrhoea as the most common manifestations are indeed gastrointestinal upset. Impacts
265 on the microbiota were investigated in dogs with acute diarrhoea and mild disease in two of the trials
266 included in the systematic review. The PCR based dysbiosis index was altered, indicating dysbiosis
267 following antimicrobial therapy with metronidazole (Rudinsky et al., 2022) but not in dogs treated with
268 amoxicillin-clavulanic acid (Werner et al., 2020). Selection for antimicrobial resistance was investigated
269 in the latter study, which documented selection of amoxicillin-resistant *Escherichia coli* (*E. coli*),
270 persisting up to 3 weeks following cessation of therapy (Grock et al., 2021).

271 When balancing the desirable against undesirable effects of antimicrobials in dogs with mild disease, the
272 panel finds that undesirable effects clearly outweigh the desirable effects, for which documentation is
273 lacking.

274

275 **Recommendations on antimicrobial use in dogs with acute diarrhoea and moderate disease**

276 **Recommendation 3**

277 In dogs with acute non-hemorrhagic diarrhoea, and moderate disease, we recommend against treatment
278 with antimicrobials.

279 *Strong recommendation, high-certainty evidence.*

280 *Level of agreement 100%*

281

282 **Recommendation 4**

283 In dogs with acute hemorrhagic diarrhoea, and moderate disease, we recommend against treatment with
284 antimicrobials.

285 *Strong recommendation, high-certainty evidence.*

286 *Level of agreement 100%*

287 **Remarks:** Dogs with laboratory values indicative of severe or overwhelming inflammation, such as
288 severe neutrophilia ($> 25 \times 10^9$), neutropenia and/or degenerative left-shift, represent an exception.

289 Clinical monitoring of dogs with moderate disease while hospitalized is imperative as some dogs will
290 experience worsening of clinical signs hours or days after initial improvement.

291 *Rationale for recommendations 3 & 4*

292 *Evidence of therapeutic effect*

293 There is high certainty evidence that antimicrobials do not confer a clinically relevant effect in dogs with
294 acute diarrhoea and moderate disease, whether or not the diarrhoea is hemorrhagic (Scahill et al., 2024),
295 2023). Based on the enquiries among dog owners and veterinarians, the risk of disease progression and
296 the duration of hospitalization are the main concerns in dogs with acute diarrhoea and moderate disease,
297 thus these are considered critical outcomes. Other outcomes deemed important in this group of dogs are
298 duration of diarrhoea and risk of mortality. The effect of antimicrobials on duration of diarrhoea in dogs
299 with acute diarrhoea is described in the prior paragraph (see evidence summary for dogs with mild
300 disease) and the same conclusion applies for dogs with moderate disease. Disease progression, duration of
301 hospitalization and mortality were investigated in the same systematic review of 232 dogs with acute
302 diarrhoea as discussed earlier (Scahill et al., 2024). Disease progression occurred in two out of 106 dogs
303 with moderate disease and hemorrhagic diarrhoea, one was described as clinically worsened and one
304 developed leukopenia. The pooled risk difference between treated and untreated dogs was 0.02, which
305 translates into a risk of 21 more dogs per 1000 dogs suffering progression of disease without
306 antimicrobials (95% CI from 70 more dogs to 30 less dogs per 1000 dogs). This risk difference was below
307 the threshold for clinical relevance predefined by panel members and veterinary practitioners, and
308 therefore considered trivial. The mean duration of hospitalization in dogs with acute diarrhoea ranged
309 from 3.59 to 3.61 days in dogs receiving antimicrobials and from 3.22 to 3.36 days in the control group.
310 When looking at the pooled mean difference between treated and untreated dogs, there was a trivial (< 24
311 hours) prolongation of time of hospitalization in dogs receiving antimicrobials by 0.37 days (95% CI
312 0.04-0.69). Likewise, for mortality there was no detectable benefit of antimicrobial therapy, and the odds
313 ratio of 1.43 (95% CI 0.24-8.54) was in favour of the untreated control group. Mortality occurred in 5 out
314 of 106 dogs with moderate disease and hemorrhagic diarrhoea, three of which were treated with
315 antimicrobials and two of which were not treated. The certainty of evidence for dogs with moderate
316 disease was high.

317 *The balance between desirable and undesirable effects*

318 When balancing the desirable against undesirable effects of antimicrobials in dogs with moderate disease,
319 the panel finds that undesirable effects clearly outweigh the desirable effects, for which documentation is
320 lacking. The readers are referred to the prior paragraph for dogs with mild disease and to Table 4 for a
321 description of the harmful effects of antimicrobials in dogs with acute diarrhoea.

322

323 **Recommendations on antimicrobial use in dogs with acute diarrhoea and severe disease**

324 **Recommendation 5**

325 In dogs with hemorrhagic and non-hemorrhagic diarrhoea, and severe disease we suggest treatment with
326 systemic antimicrobials.

327 *Conditional recommendation, low-certainty evidence*

328 *Level of agreement 100%*

329 *Rationale for recommendations 5*

330 *Evidence of therapeutic effect*

331 Dogs with severe disease constitute a minor proportion of dogs with acute diarrhoea (Singleton et al.,
332 2019) and they are not represented in any of the randomized controlled antimicrobial treatment trials
333 (Scahill et al., 2024). Observational studies in dogs with acute diarrhoea and severe disease provide data
334 on treated individuals only and baseline rates for progression of disease and mortality in untreated dogs
335 with severe disease are lacking. The overall certainty of the evidence informing the recommendation is
336 low, due to very serious indirectness of data.

337 *The balance between desirable and undesirable effects*

338 When balancing the desirable against undesirable effects of antimicrobials in dogs with severe disease,
339 the panel finds that the potential desirable effects outweigh the undesirable effects. Dogs with severe
340 disease are dogs with impaired general condition and persistent signs of systemic disease. Some dogs in
341 this group may directly present critically ill with overt signs of sepsis, while others have more subtle
342 disease, yet have not responded to - or have progressed despite - adequate fluid therapy. The panel finds
343 that discriminating between animals that will, and will not, benefit from antimicrobials in dogs with
344 severe disease is challenging, and that withholding antimicrobials may pose a risk of the disease
345 progressing to sepsis or other infectious consequences in some dogs. A beneficial effect of antimicrobials,
346 though not investigated in any trial, should be considered likely. Harmful effects of antimicrobials are

347 described in Table 4 but are considered of lesser importance to the animal's health when considering the
348 potential risk of sepsis in dogs with severe disease.

349 **Recommendation 6**

350 In dogs with severe disease, we suggest parenteral (intravenous or intramuscular) administration of
351 antimicrobials that are expected to be effective for treatment of bacterial translocation and bacteraemia or
352 sepsis. Drug choice depends on how critical the clinical status of the dog is, as well as regional AMR
353 prevalence and drug availability.

354 In dogs with non-critical illness (Table 1) living in a region with low AMR prevalence, we suggest
355 ampicillin or alternatively amoxicillin-clavulanic acid or trimethoprim/sulfonamides as first line drugs.

356 In dogs with critical illness (Table 1) or where antimicrobial resistance is more likely (e.g. based on
357 geographic trends or the patient's antimicrobial exposure history) we suggest administration of a four-
358 quadrant protocol providing gram positive, gram negative, aerobic and anaerobic coverage (Table 5).

359 Dogs with non-critical illness that do not respond to first line antimicrobials and supportive care should
360 also receive this protocol.

361 *Conditional recommendation, very low-certainty evidence/expert opinion*

362 *Level of agreement 100%*

363 *Rationale for recommendation 6*

364 *Evidence of therapeutic effect*

365 We did not identify any randomized controlled trials comparing treatment with different antimicrobials in
366 dogs with acute diarrhoea and severe disease. Some low certainty evidence can be derived from a
367 retrospective study of dogs with acute haemorrhagic diarrhoea syndrome (AHDS) in which a proportion
368 of dogs were treated with antimicrobials, the majority with intravenous ampicillin, and the prognosis was
369 favourable (Dupont et al., 2021). However, dogs included in that study were not classified into moderate
370 and severe disease and likely represented a mix of severities. The most severely ill dogs were treated with
371 a four-quadrant protocol, for the most part consisting of ampicillin and a fluoroquinolone. Indirect
372 evidence of the effect of amoxicillin clavulanic acid can be derived from the network meta-analysis
373 performed to compare the effect of metronidazole and amoxicillin clavulanic acid in dogs with mild and
374 moderate disease, in which no difference in efficacy was found (Scahill et al., 2024). However, as dogs
375 with mild and moderate disease have no benefit of treatment with antimicrobials, the value of this
376 evidence in dogs with severe disease is limited. Overall, the certainty of evidence informing the

377 recommendation on choice of treatment is very low and the recommendation is mainly based on the
378 opinion and experience of the panel members.

379 *The balance between desirable and undesirable effects*

380 In dogs with severe disease the purpose of antimicrobial administration is prevention or treatment of
381 bacterial translocation, bacteremia or sepsis, and treatment is aimed at achieving efficient systemic
382 concentrations. Parenteral administration is therefore preferred over oral therapy. De-escalation to an oral
383 equivalent can be performed once there is confidence that an oral antimicrobial will be properly absorbed.

384 When balancing benefits and harms of different antimicrobials the panel has taken into considerations the
385 limited evidence summarized above, the critical illness of the animal and the risk of developing life
386 threatening complications of infection as well as the antimicrobial spectrum and the categorisation of
387 antimicrobials for use in animals from the European Medicines Agency (EMA). EMA categorizes
388 antimicrobial drugs into four categories from D to A with D being the more prudent group ((EMA),
389 2024).

390 Based on experience from the North European countries, dogs that are not critically ill (Table 1) may
391 benefit from treatment with intravenous ampicillin (EMA cat.D) alone, or parenteral administration of
392 either amoxicillin clavulanic acid (EMA cat. C) or trimethoprim/sulphonamide (EMA cat. C).

393 For dogs with critical illness immediate administration of antimicrobial therapy with four-quadrant
394 coverage is indicated. The suggested drug combinations in Table 5 represent common combinations for
395 treatment of sepsis caused by unknown agents, it is not an exhaustive list of antimicrobial combinations
396 providing four-quadrant coverage. The panel finds that though use of fluoroquinolones (EMA cat. B)
397 should generally be restricted, their use in critically ill dogs with severe disease and acute diarrhea is
398 justified, to provide immediate coverage against gram negative *Enterobacterales*. Aminoglycosides
399 (gentamycin, amikacin) are EMA category C drugs with a gram negative spectrum similar to
400 fluoroquinolones. There is some concern over nephrotoxicity when aminoglycosides are administered to
401 animals with compromised renal blood flow, limiting their use in dogs with hypovolemia and/or reduced
402 urine production. Aminoglycosides can be administered in dogs once they are euvolemic and have
403 adequate urine production.

404 **Recommendation 7**

405 The duration of antimicrobial treatment is dependent on the treatment response and the panel suggests
406 daily assessment of animals while hospitalized. Antimicrobial therapy should not extend beyond clinical
407 resolution. For the majority of dogs, treatment of 3-7 days is likely adequate to obtain clinical resolution.

408 ***Conditional recommendation, very low-certainty evidence.***

409 ***Level of agreement 100%***

410 *Rationale for recommendation 7*

411 *Evidence of therapeutic effect*

412 There are no studies comparing the effect of short (7 days or less) vs long (greater than 7 days) duration
413 of antimicrobial treatment in dogs with acute diarrhoea. Some low certainty evidence can be derived from
414 a retrospective observational study of hospitalized dogs with AHDS, representing a mix of dogs with
415 moderate and severe disease. Of those dogs treated with antimicrobials, the majority was treated for less
416 than 7 days and up to one third of dogs were released from hospital without further antimicrobial
417 treatment (Dupont et al., 2021). Likewise, indirect evidence derived from trials in dogs with moderate
418 disease indicates that most dogs are treated for less than seven days, and that clinical resolution of disease
419 occurs prior to cessation of therapy (Scahill et al., 2024). There is currently no consensus on the optimal
420 duration of treatment in dogs with bacteraemia, or in dogs with sepsis. In people there are several RCTs
421 demonstrating that short duration (5-7 days) of antimicrobial therapy is non-inferior to long duration 10-
422 14 days) of antimicrobial therapy for gram negative bacteraemia (Runyon et al., 1991; Montravers et al.,
423 2018; Tansarli et al., 2019; Yahav et al., 2019). For suspected or established sepsis in people, recent
424 international guidelines from the Society of Critical Care Medicine (Evans et al., 2021) recommend
425 shorter over longer duration antimicrobial therapy, and daily evaluation to decide when to discontinue
426 antimicrobial therapy.

427 *The balance between desirable and undesirable effects*

428 Antimicrobial use should not be used long beyond clinical resolution to avoid harmful effects of
429 prolonged antimicrobial exposure (Table 4). It is the experience of the panel that most animals with acute
430 diarrhoea and severe disease experience resolution of disease well within 7 days of treatment, and thus
431 will not require extended treatment.

432 **Common considerations for recommendations on antimicrobial use in dogs with acute diarrhoea**

433 *Feasibility, cost and equity*

434 Recommendations are feasible and are unlikely to have important impact on equity or costs. There is a
435 cost-saving effect of not prescribing antimicrobials; however, the expenses for antimicrobial therapy vary
436 with the size of the dog and the specific product, in some cases it may constitute a relatively minor part of

437 the total cost of a veterinary consult. Antimicrobial therapy does not impact duration of hospitalization
438 and recommendations against antimicrobial therapy will not increase overall costs in hospitalized dogs.

439 *Preferences and values of dog-owners*

440 The panel considered values and preferences of veterinary prescribers and dog-owners when selecting the
441 treatments and outcomes to investigate in the systematic review (Scahill et al., 2024). Our enquiries
442 suggest that owners of dogs with acute diarrhoea have specific expectations for medication and these
443 expectations could be used to assess if therapeutic effects were clinically relevant or not (clinical effect
444 thresholds). The panel acknowledges that there may be a preference for antimicrobials when diarrhea is
445 disruptive to the owners (e.g. house soiling, waking up in the night to defecate) and that there may be
446 pressure from owners to take an approach other than watchful waiting. However, the panel believes that
447 most dog-owners would wish to avoid the cost and effort of medication if they are made aware that there
448 are no clinically relevant therapeutic effects of treatment, and in particular, if informed of the potential
449 harms of antimicrobial treatment. The panel believe that in the case of dogs with severe disease, most dog
450 owners would prefer antimicrobial treatment if informed of the potential benefits.

451 *Acceptability and implementation*

452 The panel recognizes that acceptance of a non-antimicrobial treatment strategy may vary among dog
453 owners in the European region. In some regions, guidelines will require a greater effort to implement, and
454 implementation strategies should take into account national values and preferences. Client pressure,
455 perceived or true, may pose a barrier to antimicrobial stewardship, and video animations targeting owners
456 can be downloaded, in 12 different languages, from the websites of (ENOVAT, 2024). The short video
457 animation explaining the consequences of unnecessary antimicrobial use has been tested in a population
458 of dog owners in the UK and was found to significantly impact owners' perceptions of antimicrobial use
459 (Wright, 2022)

460 **Recommendations for future research on antimicrobial use in dogs with acute diarrhea.**

461 The panel considers investigation into non-antimicrobial treatments a priority in dogs with acute
462 diarrhoea and mild or moderate disease. When investigating the effect of a given treatment one should
463 consider not only the effect on the duration of diarrhea, but also the risk of disease progression. This risk
464 is a concern of dog-owners and practitioners that was not addressed in earlier studies. Antimicrobial trials
465 are relevant in dogs with severe disease and should aim to elucidate optimal choice of drug and duration
466 of treatment. Other knowledge gaps to fill include long-term consequences of antimicrobial use and
467 diagnostic markers to identify dogs that will benefit from antimicrobial treatment.

469 Table 4. Harmful effects of commonly used antimicrobials in dogs with acute diarrhea

Harm	Description
Adverse effects	Adverse effects of antimicrobial therapy has been investigated in healthy dogs receiving metronidazole. The most common adverse effects were hyporexia, vomiting and diarrhea. Diarrhea was reported in 56% to 100% of healthy dogs following administration of metronidazole alone (Pilla et al., 2020) or in combination with enrofloxacin (Whittemore et al., 2019).
Dysbiosis	Antibiotics can lead to an alteration of the intestinal microbiota and metabolites. The severity depends on the type of antibiotic, the duration of the application, and individual factors. These changes can persist for months to years, depending on the antibiotic used and the species. Several studies in healthy dogs found that the commonly used antibiotics for canine diarrhea, tylosin and metronidazole, resulted in dysbiosis, which was present in some dogs even weeks after therapy. Moreover, typical for these antibiotics was a severe reduction in the number of <i>Clostridium hiranonis</i> (Manchester et al., 2019; Pilla et al., 2020), a bacterium that is thought to play a role in maintaining a healthy intestinal metabolism in dogs. Similarly, dysbiosis associated with metronidazole treatment in dogs with acute diarrhea was recently documented (Rudinsky et al., 2022). The alterations of the canine intestinal microbiota induced by amoxicillin or amoxicillin clavulanic acid seem to be fewer and less long lived (Gronvold et al., 2010; Espinosa-Gongora et al., 2020), and a recent study in dogs with acute diarrhea could not document dysbiosis using the PCR based dysbiosis index (Werner et al., 2020). Alterations found in other populations of dogs include reductions in microbial richness and diversity during treatment. In addition, the abundance of beneficial taxa is reduced by addition of clavulanic acid (Espinosa-Gongora et al., 2020), suggesting that clavulanic acid may broaden the impact of amoxicillin on the gut microbiota, with potential negative consequences on gut health.
Antimicrobial resistance	Selection of antimicrobial resistant bacteria is a well-documented effect of antimicrobial therapy in humans and animals, and has been documented in various populations of dogs ((Damborg et al., 2011; Espinosa-Gongora et al., 2020). In dogs with acute diarrhea, selection for antimicrobial resistance has been investigated in dogs receiving amoxicillin-clavulanic acid (Werner et al., 2020). Treatment with amoxicillin-clavulanic acid favored development of amoxicillin-resistant <i>E. coli</i> , which <i>increased</i> from 0.2% before antibiotic administration to 100% during antibiotic administration. Three weeks after discontinuation of the antibiotic, the

	percentage of amoxicillin-resistant <i>E. coli</i> was still significantly higher (10%) than in the control group (0.1%).
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470

471 Table 5. Antimicrobial combinations with four-quadrant spectrum (aerobic, anaerobic, gram positive and
472 gram negative spectrum).

Gram positives aerobes and Anaerobes	+	Gram negative Aerobes	Comments
Aminopenicillin (e.g. amoxicillin EMA D, ampicillin EMA D) or Clindamycin EMA C		Fluoroquinolone* EMA B or Aminoglycoside** EMA C (gentamycin, amikacin)	*avoid in young growing animals ** nephrotoxicity, avoid in dogs with compromised renal function or reduced renal blood flow (hypovolemia).

473 Each drug is assigned a category from the European Medical Agency (EMA): category D = Prudence,
474 category C = Caution, category B = Restrict, category A = Avoid.

475

476 **Recommendations on probiotic use in dogs with acute diarrhoea**

477 **Recommendation 8**

478 In dogs with acute diarrhoea, we do not recommend either for or against use of probiotics.

479 ***Level of agreement 100%***

480 ***Rationale for recommendations 8***

481 ***Evidence of therapeutic effect***

482 A recent systematic review conducted by this group did not identify a clinically relevant effect associated
483 with probiotic administration in dogs with acute diarrhoea (Scahill et al., 2024). The systematic review
484 included four trials of probiotic administration in dogs with acute diarrhoea, all prospective, randomized,
485 and controlled (Herstad et al., 2010; Gomez-Gallego et al., 2016; Ziese et al., 2018; Shmalberg et al., 2019).
486 The total number of dogs assessed was 149, all privately owned dogs (probiotic group = 75; placebo = 74)
487 presenting for spontaneous idiopathic acute diarrhoea. Only one study (Shmalberg et al., 2019) showed a
488 small beneficial effect of probiotics on the duration of diarrhoea (shortening of the duration of diarrhea by
489 >1 day), the effect in the other three trials was trivial, as was the effect when looking at all studies combined
490 (reduction of diarrhoea by 0.68 days). There were no clinical adverse effects or mortality reported in any
491 of the studies. Two studies showed a shift in the microbiome towards the microbiome of healthy animals

492 in the dogs receiving probiotic (Gomez-Gallego et al., 2016; Ziese et al., 2018). Risk of bias was generally
493 low, and the certainty of the combined evidence considered moderate.

494 *The balance between desirable and undesirable effects*

495 There are two major considerations leading to the panel making a non-recommendation (neither for nor
496 against). The first consideration concerns the diversity of probiotic products. Probiotics are highly diverse
497 and the biological effects are considered to be dependant not only on the specific strains, but also on the
498 dose (McFarland et al., 2018). It is unclear how the results of the systematic review apply to other probiotic
499 organisms, combinations or doses, limiting the relevance of a general recommendation for or against all
500 probiotics. The second consideration concerns the trade-offs that are closely balanced. While we could not
501 document a clinically relevant effect of treatment in dogs with acute diarrhoea, probiotics did result in what
502 was assumed to be improvements in the microbiota in two studies. However, understanding of what
503 constitutes clinically relevant beneficial changes in the gut microbiota is still limited. Probiotics are
504 considered safe in veterinary medicine. When practitioners and dog owners were questioned, we identified
505 a clear preference for probiotic prescribing among veterinary practitioners and a high degree of acceptance
506 among dog owners. However, the cost of the product, which can be considerable, and the stress of
507 medication, may not be justified. In conclusion, the panel decided not to make any recommendation
508 concerning probiotics at present, and the use of probiotics in dogs with acute diarrhoea remains a matter of
509 preference for the attending clinician and client.

510

511 **Diagnostic work up in dogs with acute diarrhoea.**

512 The following considerations on diagnostic work-up represent the professional opinion of the panel. The
513 panel has not conducted systematic reviews to inform the statements included in this section, and the
514 guidance provided is ungraded.

515

516 **Complete blood count (CBC) and Biochemistry**

517 CBC and biochemistry are indicated in dogs with acute diarrhoea and moderate or severe disease. In dogs
518 with azotemia, measurement of urine specific gravity is indicated to distinguish prerenal from renal
519 causes of azotemia.

520 *Rationale*

521 Dogs presenting with acute diarrhoea and mild disease often have self-limiting disease and do not warrant
522 extensive work up (Hubbard et al., 2007; Schwartz and Newman, 2013; Berset-Istratescu et al., 2014).
523 For dogs presenting with depressed mental status and systemic response to disease (moderate and/or
524 severe disease), a minimum database including CBC and biochemistry will help detect signs of
525 overwhelming inflammation (severe neutrophilia, neutropenia, degenerative left shift) and/or bacterial
526 sepsis (hypoglycaemia, hyperbilirubinemia), which may influence the decision to treat with an
527 antimicrobial (Purvis and Kirby, 1994; Hauptman et al., 1997). CBC and biochemistry will help assess
528 the degree of dehydration (hemoconcentration, relative hyperproteinemia, prerenal azotæmia) and detect
529 electrolyte abnormalities which may influence the amount, type and rate of fluid therapy. Lastly, it will
530 help rule out obvious metabolic or endocrine causes of acute diarrhoea.

531 **C-reactive protein (CRP)**

532 CRP may be considered in dogs with moderate or severe disease to help assess the degree of systemic
533 inflammation and monitor disease progression/regression.

534 *Rationale*

535 In dogs presenting with depressed mental status and systemic response to disease, CRP may be helpful to
536 monitor disease progression in the individual dog. It is uncertain if CRP at admission can be used for
537 antimicrobial therapy decision making, or to what extent it offers additional information compared to the
538 clinical assessment. CRP has been investigated in dogs with moderate and severe disease, more
539 specifically in dogs with parvovirus enteritis and in dogs with AHDS (McClure et al., 2013; Dupont et al.,
540 2021; Sanger et al., 2022). In dogs with AHDS, CRP correlates with clinical and laboratory scoring
541 systems (Dupont et al., 2021; Sanger et al., 2022), and concentrations decrease gradually with disease
542 regression (Sanger et al., 2022), indicating CRP might be useful as a monitoring tool. However, its benefit
543 over routine clinical monitoring remains unclear. The correlation with antimicrobial therapy has not been
544 established, and CRP did not correlate with antimicrobial therapy in a recent, prospective study in dogs
545 with AHDS (Sanger et al., 2022). In another retrospective study of dogs with AHDS (Dupont et al.,
546 2021), CRP at admission was higher in dogs that received antimicrobials compared to those receiving
547 supportive care alone. However, a causal relationship could not be established due to the retrospective
548 nature of the study, and CRP might have influenced the choice of treatment. Also, values were
549 overlapping and high CRP concentrations were also found in dogs that did not receive antimicrobials.
550 CRP did correlate to prognosis in puppies with parvovirus enteritis (McClure et al., 2013), but this was
551 not found in dogs with AHDS (Dupont et al., 2021).

552 **Testing for hypoadrenocorticism**

553 Testing for hypoadrenocorticism (e.g., adrenocorticotrophic hormone (ACTH) stimulation test/basal
554 cortisol) is indicated in dogs with acute diarrhoea presenting with either
555 depression/weakness/lethargy/collapse, and/or a history of recurrent episodes of acute diarrhoea, and/or
556 presence of laboratory abnormalities compatible with hypoadrenocorticism (Hanson et al., 2016).

557 *Rationale*

558 The clinical picture of hypoadrenocorticism ranges from mild disease to severe life-threatening vascular
559 collapse. In dogs with acute diarrhoea and mild disease a history of waxing and waning and recurrent
560 episodes may justify further work up. Typical abnormalities include hyponatemia, hyperkalemia,
561 hypercalcemia, hypoglycemia, azotemia with concurrent inability to produce concentrated urine, reverse
562 stress leukogram, lymphocytosis, eosinophilia and hypocholesterolemia (Kintzer and Peterson, 1997).

563 One should keep in mind that dogs with atypical Addison's disease do not have electrolyte abnormalities.
564 (Hauck et al., 2020). The CBC abnormalities are due to cortisol deficiency and can be found in animals
565 with both typical and atypical hypoadrenocorticism. An ACTH stimulation test may be preceded by basal
566 cortisol measurement, as values greater than 55 mmol/L (2 mcg/dL) can be used to rule out
567 hypoadrenocorticism (Lennon et al., 2007; Bovens et al., 2014). Dogs with values below 55 mmol/L
568 should undergo ACTH stimulation testing to rule the disease in or out.

569 **Diagnostic Imaging**

570 Diagnostic imaging (ultrasound/radiology) of the gastrointestinal tract is not routinely indicated in dogs
571 with acute diarrhoea. It should be considered in dogs with concomitant, non-transient, vomiting and in
572 dogs with marked or progressive abdominal pain or distension.

573 *Rationale*

574 Diagnostic imaging is generally unrewarding in dogs with diarrhoea. It is mainly relevant in dogs with
575 acute diarrhoea when gastrointestinal obstruction and/or involvement of other organ systems, such as
576 pancreatitis, is suspected (Finck et al., 2014; Mapletoft et al., 2018; Holzmann et al., 2023).

577

578 **Testing for Parvovirus**

579 Testing for canine parvovirus (CPV) enteritis (e.g., Point of Care ELISA, PCR) is indicated in young
580 dogs (< 6 months of age) with acute diarrhoea; in young dogs (<12 months of age) with acute
581 hemorrhagic diarrhoea; in unvaccinated/inadequately vaccinated dogs of any age, and should be
582 considered in dogs with neutropenia. CPV testing is also indicated whenever there is an outbreak of

583 diarrhoea in a group of (unvaccinated/inadequately vaccinated) dogs. Given the lower sensitivity of the
584 POC test for parvovirus, in cases of a negative result coupled with a strong clinical suspicion, it is
585 advisable to perform a confirmatory PCR test.

586 *Rationale*

587 Commonly, CPV infects 4–12-week-old puppies that are prone to acquiring the virus in concomitance
588 with waning maternally derived antibodies. Adults are thought to be less prone to CPV infection due to
589 the age-reduced susceptibility and presence of specific immunity induced by vaccination or previous
590 (often subclinical) infections. There are some reports of the occurrence of parvovirus in adult dogs, but
591 they are rare (Cavalli et al., 2001, Decaro et al., 2008a, Decaro et al., 2009a). The most characteristic
592 clinical form induced by CPV is represented by hemorrhagic enteritis. Leukopenia is a consistent finding,
593 with white blood cell (WBC) counts dropping below 2000–3000 cells/ μ L ($2.0\text{--}3.0 \times 10^9$ cells/L) of blood
594 (Cavalli et al., 2001, Decaro et al., 2008a, Decaro et al., 2009a).

595 **Testing for bacterial enteropathogens**

596 Testing for bacterial enteropathogens is not routinely recommended in dogs with acute diarrhoea. Faecal
597 testing can potentially be indicated in dogs with severe disease that are of increased risk of pathogen
598 transmission (e.g., fed a raw diet) or when several individuals in a household, including dog owners, or in
599 the local region show similar clinical signs. Testing for *Clostridium difficile* (PCR combined with ELISA
600 for toxin A/B), *Campylobacter jejuni/coli* (PCR or culture), and *Salmonella* spp. (PCR or culture) could
601 be considered in these cases. However, antimicrobial treatment is not recommended beyond the resolution
602 of clinical signs even when test results prove positive for enteropathogens. Bacterial culture of blood,
603 abdominal fluid or lymph node aspirates should be considered if sepsis or bacteraemia/bacterial
604 translocation is suspected.

605 *Rationale*

606 Canine acute diarrhoea is self-limiting, and faecal testing is thus unlikely to change treatment
607 recommendations (Cave et al., 2002). Dogs that are fed a raw diet are at increased risk of transmitting
608 antimicrobial resistant bacteria, as well as *Salmonella* spp and *Campylobacter* spp (Viegas et al., 2020).
609 Testing in these animals can be considered to elucidate the zoonotic risk, a positive test result is not an
610 indication to treat with antimicrobials. *Clostridium perfringens* is part of the microbiota in healthy dogs
611 (Werner et al., 2021), and although strains encoding for NetF-toxin might play a role in acute
612 haemorrhagic diarrhoea syndrome, testing is still not recommended since a positive result does not
613 change treatment recommendations (Sindern et al., 2019). If testing for *C. difficile* is performed, an

614 ELISA for toxin A/B should be included as this could be part of the aetiology in a small number of
615 individuals (Rainha et al., 2022). Haemolytic *E. coli* is found in the gastrointestinal microbiota in healthy
616 dogs, as well as in dogs with diarrhoea, and testing is not recommended (Werner et al., 2021).

617 **Fecal flotation and testing for *Giardia***

618 Testing for *Giardia* should be considered in young dogs with acute diarrhoea and is particularly indicated
619 in those with non-self-limiting or relapsing disease.

620 *Rationale*

621 Protozoa or parasites are infrequently the primary cause of diarrhoea and might be a coincidental finding
622 with a prevalence of 7-17 % (Drake et al., 2022) in healthy dogs and somewhat higher prevalence in
623 hunting or shelter dogs (Uiterwijk et al., 2019). Most cases are not associated with clinical signs.
624 Nevertheless, parasites are thought to lower the threshold for diarrhoea caused by other factors and are
625 typically treated in dogs with clinical disease. qPCR is by far the most sensitive test for *Giardia* detection
626 and might be useful to assess the zoonotic risk, but may not reflect a clinically relevant protozoal load
627 causing diarrhea (passage of ingested cysts through the intestine). Fecal antigen testing and fecal flotation
628 are probably the most widely used tests in veterinary practice for *Giardia* detection. Antigen testing is
629 more sensitive than fecal flotation (Uiterwijk et al., 2018) and the combined use of both methods
630 improves detection (Drake et al., 2022).

631

632 **References**

633 (EMA), E.M.A., 2024. Categorisation of antibiotics used in animals promotes responsible use to protect
634 public and animal health. [https://www.ema.europa.eu/en/news/categorisation-antibiotics-
635 used-animals-promotes-responsible-use-protect-public-and-animal-health](https://www.ema.europa.eu/en/news/categorisation-antibiotics-used-animals-promotes-responsible-use-protect-public-and-animal-health) (Accessed 23.
636 February, 2024).

637

638 Alonso-Coello, P., Oxman, A.D., Moberg, J., Brignardello-Petersen, R., Akl, E.A., Davoli, M., Treweek, S.,
639 Mustafa, R.A., Vandvik, P.O., Meerpohl, J., et al., 2016. GRADE Evidence to Decision (EtD)
640 frameworks: a systematic and transparent approach to making well informed healthcare
641 choices. 2: Clinical practice guidelines. *BMJ* 353, i2089.

642

643 Berset-Istratescu, C.M., Glardon, O.J., Magouras, I., Frey, C.F., Gobeli, S., Burgener, I.A., 2014. Follow-up
644 of 100 dogs with acute diarrhoea in a primary care practice. *Vet J* 199, 188-190.

645

646 Bovens, C., Tennant, K., Reeve, J., Murphy, K.F., 2014. Basal serum cortisol concentration as a screening
647 test for hypoadrenocorticism in dogs. *J Vet Intern Med* 28, 1541-1545.

648
649 Cave, N.J., Marks, S.L., Kass, P.H., Melli, A.C., Brophy, M.A., 2002. Evaluation of a routine diagnostic fecal
650 panel for dogs with diarrhea. *J Am Vet Med Assoc* 221, 52-59.

651
652 Damborg, P., Gaustad, I.B., Olsen, J.E., Guardabassi, L., 2011. Selection of CMY-2 producing *Escherichia*
653 *coli* in the faecal flora of dogs treated with cephalexin. *Vet Microbiol* 151, 404-408.

654
655 De Briyne, N., Atkinson, J., Pokludova, L., Borriello, S.P., Price, S., 2013. Factors influencing antibiotic
656 prescribing habits and use of sensitivity testing amongst veterinarians in Europe. *Vet Rec* 173,
657 475.

658
659 Drake, J., Sweet, S., Baxendale, K., Hegarty, E., Horr, S., Friis, H., Goddu, T., Ryan, W.G., von Samson-
660 Himmelstjerna, G., 2022. Detection of *Giardia* and helminths in Western Europe at local K9
661 (canine) sites (DOGWALKS Study). *Parasit Vectors* 15, 311.

662
663 Dupont, N., Jessen, L.R., Moberg, F., Zyskind, N., Lorentzen, C., Bjornvad, C.R., 2021. A retrospective
664 study of 237 dogs hospitalized with suspected acute hemorrhagic diarrhea syndrome: Disease
665 severity, treatment, and outcome. *J Vet Intern Med* 35, 867-877.

666
667 ENOVAT, 2024. The European Network for Optimization of Antimicrobial Therapy. <https://enovat.eu/>
668 (Accessed 23. february, 2024).

669
670 Espinosa-Gongora, C., Jessen, L.R., Kieler, I.N., Damborg, P., Bjornvad, C.R., Gudeta, D.D., Pires Dos
671 Santos, T., Sablier-Gallis, F., Sayah-Jeanne, S., Corbel, T., et al., 2020. Impact of oral amoxicillin
672 and amoxicillin/clavulanic acid treatment on bacterial diversity and beta-lactam resistance in the
673 canine faecal microbiota. *J Antimicrob Chemother* 75, 351-361.

674
675 Evans, L., Rhodes, A., Alhazzani, W., Antonelli, M., Coopersmith, C.M., French, C., Machado, F.R.,
676 McIntyre, L., Ostermann, M., Prescott, H.C., et al., 2021. Surviving sepsis campaign: international
677 guidelines for management of sepsis and septic shock 2021. *Intensive Care Med* 47, 1181-1247.

678
679 Finck, C., D'Anjou, M.A., Alexander, K., Specchi, S., Beauchamp, G., 2014. Radiographic diagnosis of
680 mechanical obstruction in dogs based on relative small intestinal external diameters. *Vet Radiol*
681 *Ultrasound* 55, 472-479.

682
683 Gomez-Gallego, C., Junnila, J., Mannikko, S., Hameenoja, P., Valtonen, E., Salminen, S., Beasley, S., 2016.
684 A canine-specific probiotic product in treating acute or intermittent diarrhea in dogs: A double-
685 blind placebo-controlled efficacy study. *Vet Microbiol* 197, 122-128.

686
687 Grock, A., Jordan, J., Zaver, F., Colmers-Gray, I.N., Krishnan, K., Chan, T., Thoma, B., Alexander, C.,
688 Alkhalifah, M., Almehli, A.S., et al., 2021. The revised Approved Instructional Resources score:

689 An improved quality evaluation tool for online educational resources. *AEM Educ Train* 5,
690 e10601.

691

692 Gronvold, A.M., L'Abée-Lund, T.M., Sorum, H., Skancke, E., Yannarell, A.C., Mackie, R.I., 2010. Changes in
693 fecal microbiota of healthy dogs administered amoxicillin. *FEMS Microbiol Ecol* 71, 313-326.

694

695 Guyatt, G.H., Oxman, A.D., Vist, G.E., Kunz, R., Falck-Ytter, Y., Alonso-Coello, P., Schunemann, H.J.,
696 Group, G.W., 2008. GRADE: an emerging consensus on rating quality of evidence and strength of
697 recommendations. *BMJ* 336, 924-926.

698

699 Hanson, J.M., Tengvall, K., Bonnett, B.N., Hedhammar, A., 2016. Naturally Occurring Adrenocortical
700 Insufficiency--An Epidemiological Study Based on a Swedish-Insured Dog Population of 525,028
701 Dogs. *J Vet Intern Med* 30, 76-84.

702

703 Hauck, C., Schmitz, S.S., Burgener, I.A., Wehner, A., Neiger, R., Kohn, B., Rieker, T., Reese, S., Unterer, S.,
704 2020. Prevalence and characterization of hypoadrenocorticism in dogs with signs of chronic
705 gastrointestinal disease: A multicenter study. *J Vet Intern Med* 34, 1399-1405.

706

707 Hauptman, J.G., Walshaw, R., Olivier, N.B., 1997. Evaluation of the sensitivity and specificity of
708 diagnostic criteria for sepsis in dogs. *Vet Surg* 26, 393-397.

709

710 Herstad, H.K., Nesheim, B.B., L'Abée-Lund, T., Larsen, S., Skancke, E., 2010. Effects of a probiotic
711 intervention in acute canine gastroenteritis--a controlled clinical trial. *J Small Anim Pract* 51, 34-
712 38.

713

714 Holzmann, B., Werner, M., Unterer, S., Dorfelt, R., 2023. Utility of diagnostic tests in vomiting dogs
715 presented to an internal medicine emergency service. *Front Vet Sci* 10, 1063080.

716

717 Hubbard, K., Skelly, B.J., McKelvie, J., Wood, J.L., 2007. Risk of vomiting and diarrhoea in dogs. *Vet Rec*
718 161, 755-757.

719

720 Israeloff, J.V. 2009. Vergleich von Therapieformen der idiopathischen hämorrhagischen Gastroenteritis
721 (HGE) beim Hund. . Dissertation, PhD, Veterinärmedizinischen Universität Wien

722

723 Jones, P.H., Dawson, S., Gaskell, R.M., Coyne, K.P., Tierney, A., Setzkorn, C., Radford, A.D., Noble, P.J.,
724 2014. Surveillance of diarrhoea in small animal practice through the Small Animal Veterinary
725 Surveillance Network (SAVSNET). *Vet J* 201, 412-418.

726

727 Kintzer, P.P., Peterson, M.E., 1997. Primary and secondary canine hypoadrenocorticism. *Vet Clin North*
728 *Am Small Anim Pract* 27, 349-357.

729

730 Langlois, D.K., Koenigshof, A.M., Mani, R., 2020. Metronidazole treatment of acute diarrhea in dogs: A
731 randomized double blinded placebo-controlled clinical trial. *J Vet Intern Med* 34, 98-104.

732

733 Lennon, E.M., Boyle, T.E., Hutchins, R.G., Friedenthal, A., Correa, M.T., Bissett, S.A., Moses, L.S., Papich,
734 M.G., Birkenheuer, A.J., 2007. Use of basal serum or plasma cortisol concentrations to rule out a
735 diagnosis of hypoadrenocorticism in dogs: 123 cases (2000-2005). *J Am Vet Med Assoc* 231, 413-
736 416.

737

738 Lutz, B., Lehner, C., Schmitt, K., Willi, B., Schupbach, G., Mevissen, M., Peter, R., Muntener, C., Naegeli,
739 H., Schuller, S., 2020. Antimicrobial prescriptions and adherence to prudent use guidelines for
740 selected canine diseases in Switzerland in 2016. *Vet Rec Open* 7, e000370.

741

742 Manchester, A.C., Webb, C.B., Blake, A.B., Sarwar, F., Lidbury, J.A., Steiner, J.M., Suchodolski, J.S., 2019.
743 Long-term impact of tylosin on fecal microbiota and fecal bile acids of healthy dogs. *J Vet Intern*
744 *Med* 33, 2605-2617.

745

746 Mapletoft, E.K., Allenspach, K., Lamb, C.R., 2018. How useful is abdominal ultrasonography in dogs with
747 diarrhoea? *J Small Anim Pract* 59, 32-37.

748

749 McClure, V., van Schoor, M., Thompson, P.N., Kjelgaard-Hansen, M., Goddard, A., 2013. Evaluation of
750 the use of serum C-reactive protein concentration to predict outcome in puppies infected with
751 canine parvovirus. *J Am Vet Med Assoc* 243, 361-366.

752

753 McFarland, L.V., Evans, C.T., Goldstein, E.J.C., 2018. Strain-Specificity and Disease-Specificity of Probiotic
754 Efficacy: A Systematic Review and Meta-Analysis. *Front Med (Lausanne)* 5, 124.

755

756 Montravers, P., Tubach, F., Lescot, T., Veber, B., Esposito-Farese, M., Seguin, P., Paugam, C., Lepape, A.,
757 Meistelman, C., Cousson, J., et al., 2018. Short-course antibiotic therapy for critically ill patients
758 treated for postoperative intra-abdominal infection: the DURAPOP randomised clinical trial.
759 *Intensive Care Med* 44, 300-310.

760

761 Mortier, F., Strohmeier, K., Hartmann, K., Unterer, S., 2015. Acute haemorrhagic diarrhoea syndrome in
762 dogs: 108 cases. *Vet Rec* 176, 627.

763

764 Pilla, R., Gaschen, F.P., Barr, J.W., Olson, E., Honneffer, J., Guard, B.C., Blake, A.B., Villanueva, D.,
765 Khattab, M.R., AlShawaqfeh, M.K., et al., 2020. Effects of metronidazole on the fecal
766 microbiome and metabolome in healthy dogs. *J Vet Intern Med* 34, 1853-1866.

767

768 Purvis, D., Kirby, R., 1994. Systemic inflammatory response syndrome: septic shock. *Vet Clin North Am*
769 *Small Anim Pract* 24, 1225-1247.

770

771 Rainha, K., Lins, D., Ferreira, R.F., Costa, C.L., Penna, B., Endres, B.T., Garey, K.W., Domingues, R.,
772 Ferreira, E.O., 2022. Colitis caused by Clostridioides difficile infection in a domestic dog: A case
773 report. Anaerobe 73, 102511.

774
775 Rudinsky, A.J., Parker, V.J., Winston, J., Cooper, E., Mathie, T., Howard, J.P., Bremer, C.A., Yaxley, P.,
776 Marsh, A., Laxalde, J., et al., 2022. Randomized controlled trial demonstrates nutritional
777 management is superior to metronidazole for treatment of acute colitis in dogs. J Am Vet Med
778 Assoc 260, S23-S32.

779
780 Runyon, B.A., McHutchison, J.G., Antillon, M.R., Akriviadis, E.A., Montano, A.A., 1991. Short-course
781 versus long-course antibiotic treatment of spontaneous bacterial peritonitis. A randomized
782 controlled study of 100 patients. Gastroenterology 100, 1737-1742.

783
784 Sanger, F., Unterer, S., Werner, M., Dorfelt, R., 2022. C-reactive protein as a tool for monitoring
785 response to treatment in dogs with acute hemorrhagic diarrhea syndrome. Front Vet Sci 9,
786 1019700.

787
788 Scahill, K., Jessen, L.R., Prior, C., Singleton, D., Foroutan, F., Ferran, A.A., Arenas, C., Bjornvad, C.R., Lavy,
789 E., Allerton, F., et al., 2024. Efficacy of antimicrobial and nutraceutical treatment for canine
790 acute diarrhoea: A systematic review and meta-analysis for European Network for Optimization
791 of Antimicrobial Therapy (ENOVAT) guidelines. Vet J 303, 106054.

792
793 Schwartz, S., Newman, B., 2013. Vomiting and diarrhoea in dogs. Vet Rec 172, 136.

794
795 Shmalberg, J., Montalbano, C., Morelli, G., Buckley, G.J., 2019. A Randomized Double Blinded Placebo-
796 Controlled Clinical Trial of a Probiotic or Metronidazole for Acute Canine Diarrhea. Front Vet Sci
797 6, 163.

798
799 Sindern, N., Suchodolski, J.S., Leutenegger, C.M., Mehdizadeh Gohari, I., Prescott, J.F., Proksch, A.L.,
800 Mueller, R.S., Busch, K., Unterer, S., 2019. Prevalence of Clostridium perfringens netE and netF
801 toxin genes in the feces of dogs with acute hemorrhagic diarrhea syndrome. J Vet Intern Med
802 33, 100-105.

803
804 Singleton, D.A., Noble, P.J.M., Sanchez-Vizcaino, F., Dawson, S., Pinchbeck, G.L., Williams, N.J., Radford,
805 A.D., Jones, P.H., 2019. Pharmaceutical Prescription in Canine Acute Diarrhoea: A Longitudinal
806 Electronic Health Record Analysis of First Opinion Veterinary Practices. Front Vet Sci 6, 218.

807
808 Tansarli, G.S., Andreatos, N., Pliakos, E.E., Mylonakis, E., 2019. A Systematic Review and Meta-analysis of
809 Antibiotic Treatment Duration for Bacteremia Due to Enterobacteriaceae. Antimicrob Agents
810 Chemother 63.

811

812 Uiterwijk, M., Nijse, R., Kooyman, F.N.J., Wagenaar, J.A., Mughini-Gras, L., Koop, G., Ploeger, H.W.,
813 2018. Comparing four diagnostic tests for *Giardia duodenalis* in dogs using latent class analysis.
814 *Parasit Vectors* 11, 439.

815

816 Uiterwijk, M., Nijse, R., Kooyman, F.N.J., Wagenaar, J.A., Mughini-Gras, L., Ploeger, H.W., 2019. Host
817 factors associated with *Giardia duodenalis* infection in dogs across multiple diagnostic tests.
818 *Parasit Vectors* 12, 556.

819

820 Unterer, S., Strohmeyer, K., Kruse, B.D., Sauter-Louis, C., Hartmann, K., 2011. Treatment of aseptic dogs
821 with hemorrhagic gastroenteritis with amoxicillin/clavulanic acid: a prospective blinded study. *J*
822 *Vet Intern Med* 25, 973-979.

823

824 Viegas, F.M., Ramos, C.P., Xavier, R.G.C., Lopes, E.O., Junior, C.A.O., Bagno, R.M., Diniz, A.N., Lobato,
825 F.C.F., Silva, R.O.S., 2020. Fecal shedding of *Salmonella* spp., *Clostridium perfringens*, and
826 *Clostridioides difficile* in dogs fed raw meat-based diets in Brazil and their owners' motivation.
827 *PLoS One* 15, e0231275.

828

829 Werner, M., Suchodolski, J.S., Lidbury, J.A., Steiner, J.M., Hartmann, K., Unterer, S., 2021. Diagnostic
830 value of fecal cultures in dogs with chronic diarrhea. *J Vet Intern Med* 35, 199-208.

831

832 Werner, M., Suchodolski, J.S., Straubinger, R.K., Wolf, G., Steiner, J.M., Lidbury, J.A., Neuerer, F.,
833 Hartmann, K., Unterer, S., 2020. Effect of amoxicillin-clavulanic acid on clinical scores, intestinal
834 microbiome, and amoxicillin-resistant *Escherichia coli* in dogs with uncomplicated acute
835 diarrhea. *J Vet Intern Med* 34, 1166-1176.

836

837 Whittemore, J.C., Moyers, T.D., Price, J.M., 2019. Randomized, controlled, crossover trial of prevention
838 of antibiotic-induced gastrointestinal signs using a synbiotic mixture in healthy research dogs. *J*
839 *Vet Intern Med* 33, 1619-1626.

840

841 WHO, 2024. World Health Organization. [https://www.who.int/health-topics/antimicrobial-](https://www.who.int/health-topics/antimicrobial-resistance2024)
842 [resistance2024](https://www.who.int/health-topics/antimicrobial-resistance2024)).

843

844 Wright, E., Pflieger, S., Gajanayake, I., Gemmill, T., Jessen, Lisbeth Rem, Sørensen, Tina Møller, Battersby,
845 I., West, E., Atkinson, L., Mosher, M., Rutland, C., Singleton, D., Tompson, A. & Allerton, F., 2022.
846 The impact of a short, animated film on owner attitudes towards antimicrobial use in companion
847 animals—a randomised controlled trial. , In: *Journal of veterinary internal medicine / American*
848 *College of Veterinary Internal Medicine*. 36 , 6, s. 2502 1 s.

849 .

850

851 Yahav, D., Franceschini, E., Koppel, F., Turjeman, A., Babich, T., Bitterman, R., Neuberger, A., Ghanem-
852 Zoubi, N., Santoro, A., Eliakim-Raz, N., et al., 2019. Seven Versus 14 Days of Antibiotic Therapy

853 for Uncomplicated Gram-negative Bacteremia: A Noninferiority Randomized Controlled Trial.
854 Clin Infect Dis 69, 1091-1098.

855
856 Zeng, L., Brignardello-Petersen, R., Hultcrantz, M., Mustafa, R.A., Murad, M.H., Iorio, A., Traversy, G.,
857 Akl, E.A., Mayer, M., Schunemann, H.J., et al., 2022. GRADE Guidance 34: update on rating
858 imprecision using a minimally contextualized approach. J Clin Epidemiol 150, 216-224.

859
860 Ziese, A.L., Suchodolski, J.S., Hartmann, K., Busch, K., Anderson, A., Sarwar, F., Sindern, N., Unterer, S.,
861 2018. Effect of probiotic treatment on the clinical course, intestinal microbiome, and toxigenic
862 Clostridium perfringens in dogs with acute hemorrhagic diarrhea. PLoS One 13, e0204691.

863

864

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